


**IEC
TSKgel BioAssist SERIES**
APPLICATIONS WITH TSKgel BioAssist ANION EXCHANGE COLUMNS
Comparison of Dynamic Binding Capacity

Table 2 shows typical dynamic binding capacities on Bio-Assist Q relative to competitive products.

TABLE II
COMPARISON OF DYNAMIC BINDING CAPACITIES

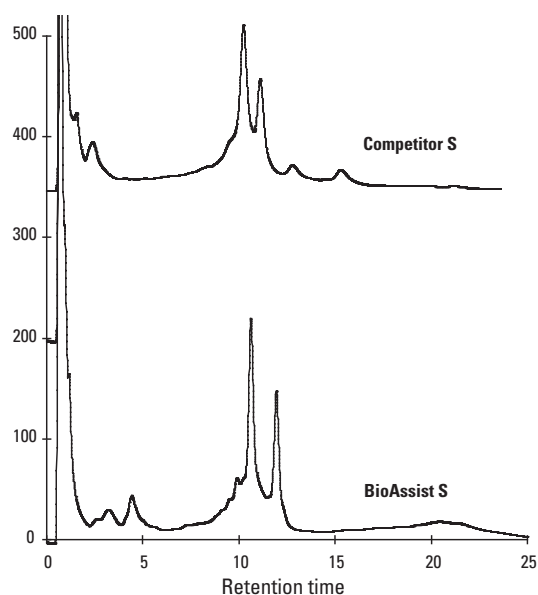
Protein	Binding capacity (mg/mL)			
	BioAssist Q	SuperQ -5PW	Conv. Q type prod. A	Conv. Q type prod. B
Thyroglobulin	77.4	22.9	20.2	1.8
Monoclonal IgG ₁	57.8	43.3	46.7	47.7
Human Serum Albumin	83.1	78.9	48.2	48.8
Trypsin Inhibitor	84.3	92.8	51.8	57.8

Columns: TSKgel BioAssist Q and TSKgel SuperQ-5PW (4.6 mm ID x 1 cm);
Conventional Q type product A and B (4.6 mm ID x 1 cm)

Solvent: 20 mmol/L Tris-HCl buffer, pH 8.0;

Flow rate: 0.38 mL/min; Det.: UV @ 280 nm

*Capacity was determined at 10% height of the breakthrough curve; UV 280 nm.

FIGURE 12
BROMELAIN ANALYSIS ON TSKgel BioAssist S & COMPETITOR S COLUMNS


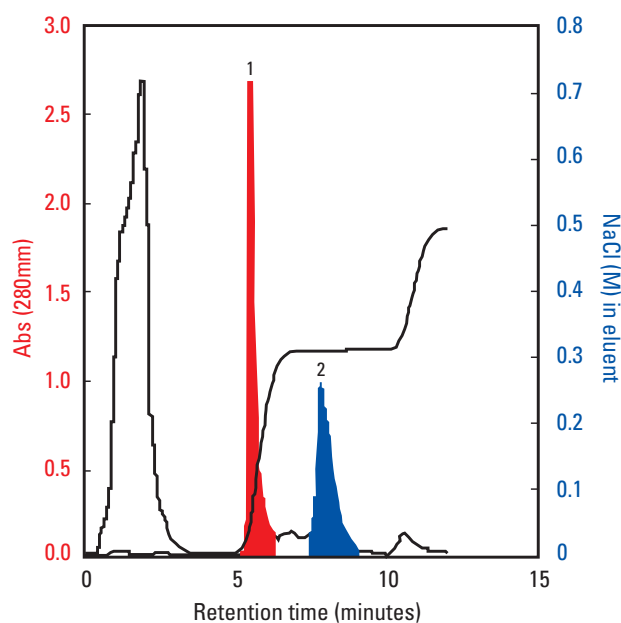
Columns: TSKgel BioAssist S, 4.6 mm ID x 5 cm L, PEEK
Competitor S 5 mm ID x 5 cm L; Elution: 20 min (TSKgel) or 30 min (Competitor S) linear gradient of NaCl from 0 to 0.5 mol/L in 20 mmol/L sodium phosphate buffer, pH 7.0; Flow rate: 0.8 mL/min for TSKgel; 1.0 mL/min for Competitor S
Detection: UV @ 280 nm; Temperature: 25°C;
Sample: crude bromelain (C4882, Sigma), 1 mg in 100 μ L

APPLICATIONS WITH TSKgel BioAssist CATION EXCHANGE COLUMNS
Bromelain Analysis on TSKgel BioAssist S and Competitor S Column

Figure 12 shows the analysis of bromelain, a proteolytic enzyme that is used as a nutritional supplement. Bromelain is a basic glycoprotein with a MW of 33 kDa and a pI of 9.55.

Analysis of IgM

IgM is known to possess unique and beneficial characteristics relative to other immunoglobulin classes; it is a large molecule comprised of five IgG subunits, resulting in a relatively unstable and difficult to purify protein. Unlike single chain antibodies, IgM cannot be purified by Protein A (affinity material commonly used for its high binding capacity and excellent selectivity for antibodies) due to steric hindrance. Alternative affinity methods have been developed with thiophilic adsorbents but these methods often result in low binding capacity. An alternative purification method of IgM by ion exchange chromatography using a TSKgel BioAssist S column was developed. Figure 13 shows the baseline separation of IgM from other contaminants using a 0.3 mol/L NaCl step gradient after elution of albumin.

FIGURE 13
ANALYSIS OF IgM


Column: TSKgel BioAssist S, 7 μ m, 4.6 mm ID x 5 cm L;
Mobile phase: 20 mmol/L sodium phosphate buffer, pH 6.0;
Gradient: 0 mol/L - 0.3 mol/L NaCl (5 min), 0.3 mol/L - 0.5 mol/L NaCl (10 min);
Flow rate: 1 mL/min; Detection: UV @ 280 nm; Sample: 500 μ L of 9.5 mg/mL IgM in mouse ascites fluid; shaded peaks represent albumin and IgM respectively